

Drug Development

Jeffrey Hasday (ahasday@som.umaryland.edu): The Hasday lab has focused on how febrile-range hyperthermia and hypothermia modify biological processes relevant to disease pathogenesis with emphasis on acute lung injury/ARDS and fibrosis. Using approaches that span structural biology, gene and protein expression, cell culture, animal models and human trials, the Hasday laboratory has shown that hyperthermia worsens and hypothermia improves lung injury by modifying endothelial permeability, neutrophil recruitment, epithelial injury, and cytokine and heat shock protein expression. The p38 MAP kinase pathway appears to be a major contributor to the temperature-dependence of endothelial barrier function and expression of pro-inflammatory cytokines. The Hasday laboratory is currently has three areas of research: (1) the DoD-funded Cooling to Help Injured Lungs (CHILL) randomized clinical trial of mild hypothermia plus neuromuscular blockade vs. standard temperature management in patients with moderate to severe ARDS, a 14-center trial for which we serve as both the Data Coordinating Center and Clinical Coordinating Center; (2) expansion of our exciting data showing that the structure and function of p38alpha, the proinflammatory p38 family member, but not p38beta is temperature-dependent in the 33° to 39°C range; and (3) the computer-assisted design of a novel class of substrate- and function-selective inhibitors of p38alpha for treatment of acute lung injury; one of these novel drugs has just completed Phase 1 studies and will begin Phase 2 studies in the near future. Dr. Hasday also directs the University of Maryland Cytokine Core Laboratory (www.cytokines.com).

Highlighted Publications:

1. Shah NG, Tulapurkar ME, Ramarathnam A, Brophy A, Martinez R 3rd, Hom K, Hodges, T, Samadani R, Singh IS, MacKerell AD Jr, Shapiro P, Hasday JD. Novel Noncatalytic Substrate-Selective p38 α -Specific MAPK Inhibitors with Endothelial-Stabilizing and Anti-Inflammatory Activity. *J Immunol.* 2017; 198(8):3296-3306. Pubmed PMID: 28298524.
2. Slack DF, Corwin DS, Shah NG, Shanholtz CB, Verceles AC, Netzer G, Jones KM, Brown CH, Terrin ML, Hasday JD. Pilot Feasibility Study of Therapeutic Hypothermia for Moderate to Severe Acute Respiratory Distress Syndrome. *Crit Care Med.* 2017 45:1152-59;PubMed PMID: [28406814](https://pubmed.ncbi.nlm.nih.gov/28406814/).
3. Tulapurkar ME, Ramarathnam A, Hasday JD, Singh IS. Bacterial lipopolysaccharide augments febrile-range hyperthermia-induced heat shock protein 70 expression and extracellular release in human THP1 cells. *PLoS One.* 2015;10(2):e0118010. PubMed PMID: [25659128](https://pubmed.ncbi.nlm.nih.gov/25659128/); PubMed Central PMCID: [PMC4320107](https://pubmed.ncbi.nlm.nih.gov/PMC4320107/).
4. Gupta A, Cooper ZA, Tulapurkar ME, Potla R, Maity T, Hasday JD, Singh IS. Toll-like receptor agonists and febrile range hyperthermia synergize to induce heat shock protein 70 expression and extracellular release. *J Biol Chem.* 2013 Jan 25;288(4):2756-66. PubMed PMID: [23212905](https://pubmed.ncbi.nlm.nih.gov/23212905/); PubMed Central PMCID: [PMC3554941](https://pubmed.ncbi.nlm.nih.gov/PMC3554941/).
5. Tulapurkar ME, Almutairy EA, Shah NG, He JR, Puche AC, Shapiro P, Singh IS, Hasday JD. Febrile-range hyperthermia modifies endothelial and neutrophilic functions to promote extravasation. *Am J Respir Cell Mol Biol.* 2012 Jun;46(6):807-14. PubMed PMID: [22281986](https://pubmed.ncbi.nlm.nih.gov/22281986/); PubMed Central PMCID: [PMC3380289](https://pubmed.ncbi.nlm.nih.gov/PMC3380289/).
6. Shah NG, Tulapurkar ME, Damarla M, Singh IS, Goldblum SE, Shapiro P, Hasday JD. Febrile-range hyperthermia augments reversible TNF- α -induced hyperpermeability in human microvascular lung endothelial cells. *Int J Hyperthermia.* 2012;28(7):627-35. PubMed PMID: [22834633](https://pubmed.ncbi.nlm.nih.gov/22834633/).
7. Deredge, D., Wintrode, P., Tulapiurkar, M. E., Nagarsekar, A., Zhang, Y., Weber, D. J., Shapiro, P., and Hasday, J. D. (2019) A temperature-dependent conformational shift in p38□

MAP kinase substrate binding region associated with changes in substrate phosphorylation profile. *J. Biol. Chem.* 2019;294:1264-37. PubMed PMID 31073086.

Links:

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PubMed publications:

<https://www.ncbi.nlm.nih.gov/myncbi/browse/collection/40776367/?sort=date&direction=ascending>

Paul Shapiro (pshapiro@rx.umaryland.edu): The primary objective of the Shapiro laboratory is to identify novel small molecular weight compounds that can selectively disrupt the interactions between protein kinases and substrate proteins to afford a more specific manipulation of signaling networks involved in disease. The experimental approach integrates computational chemistry with structural and functional cell biology. The laboratory has focused on identifying compounds that interact with unique docking sites on the extracellular signal regulated kinases (ERKs) and p38 and disrupt phosphorylation of select protein substrates. The ERK inhibitors are being tested in models of cancer and the airway smooth muscle hypertrophy that occurs in asthma. The p38 inhibitors developed in collaboration with Dr. Hasday, are being developed as anti-inflammatory agents and tested in models of acute lung injury.

Highlighted Publications:

1. Chad N. Hancock, Alba Macias, Eun Kyoung Lee, Su Yeon Yu, Alexander D. MacKerell, Jr., and Paul Shapiro (2005) Identification of novel extracellular signal-regulated kinase (ERK) docking domain inhibitors. *Journal of Medicinal Chemistry*, 48:4586-4595. PMID: 15999996
2. Fengming Chen, Chad N. Hancock, Alba T. Macias, Joseph Joh, Kimberly Still, Shijun Zhong, Alexander D. MacKerell, and Paul Shapiro (2006) Characterization of ATP-independent ERK inhibitors identified through in silico analysis of the active ERK2 structure. *Bioorganic Medicinal Chemistry Letters*, 16:6281-6287. PMID: 17000106
3. Shah NG, Tulapurkar ME, Ramarathnam A, Brophy A, Martinez R 3rd, Hom K, Hodges T, Samadani R, Singh IS, MacKerell AD Jr, Shapiro P, Hasday JD. (2017) Novel Noncatalytic Substrate-Selective p38 α -Specific MAPK Inhibitors with Endothelial-Stabilizing and Anti-Inflammatory Activity. *J Immunol.* 198: 3296–3306. PMID: 28298524
4. Kira G. Hartman, Michele I. Vitolo, Adam D. Pierce, Jennifer M. Fox, Paul Shapiro, Stuart S. Martin, Paul T. Wilder and David J. Weber. (2014) Complex formation between S100B and the p90 ribosomal S6 kinase (RSK) in malignant melanoma is Ca²⁺-dependent and inhibits ERK-mediated phosphorylation of RSK. *J. Biol. Chem.* 289(18):12886-95. PMID:24627490
5. Samadani R, Zhang J, Brophy A, Oashi T, Priyakumar UD, Raman EP, St John FJ, Jung KY, Fletcher S, Pozharski E, MacKerell AD, Shapiro PS (2015). Small Molecule Inhibitors of ERK-mediated Immediate Early Gene Expression and Proliferation of Melanoma Cells Expressing Mutated BRAf. *Biochem J.* Volume 467 (part 3), p425-438. PMID: 25695333

Links:

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Konstantin Birukov (kbirukov@som.umaryland.edu): Dr. Birukov's research is aimed at better understanding of molecular events driving onset and resolution of acute lung injury and development of new therapies to mitigate pathologic signaling leading to ARDS. The studies cover several areas, including: a) role of circulating danger associated molecular patterns (DAMPs) as biomarkers and pathogenic factors augmenting ARDS; b) synergy between mechanical stretch,

tissue stiffness and bacterial pathogens in propagation of lung injury and inflammation; c) control of endothelial function by mechanical forces; d) role of oxidized phospholipids in lung pathobiology and development of new phospholipid-based therapeutics. His laboratory uses advanced biophysical and imaging methods, endothelial cell culture models of mechanical stress, and animal models of lung injury to understand the autoregulatory cascades providing recovery and resolution of acute lung injury. He developed a new area of research addressing novel, barrier-protective and anti-inflammatory properties of oxidized phospholipids and proposed a new group of synthetic phospholipase resistant lipid mediators for future treatment of lung injury, inflammation and vascular barrier dysfunction.

Highlighted Publications:

1. Everitt ML, Boegner DJ, Birukov KG, White IM. Sample-to-answer diagnostic system for the detection of circulating histones in whole blood. *ACS Sens.* 2021
2. Wyman AE, Nguyen TTT, Karki P, Tulapurkar ME, Zhang CO, Kim J, Feng TG, Dabo AJ, Todd NW, Luzina IG, Geraghty P, Foronjy RF, Hasday JD, Birukova AA, Atamas SP, Birukov KG. Sirt7 deficiency suppresses inflammation, induces EndoMT, and increases vascular permeability in primary pulmonary endothelial cells. *Sci Rep.* 2020;10:12497
3. Kim J, Nguyen TTT, Li Y, Zhang CO, Cha B, Ke Y, Mazzeffi MA, Tanaka KA, Birukova AA, Birukov KG. Contrasting effects of stored allogeneic red blood cells and their supernatants on permeability and inflammatory responses in human pulmonary endothelial cells. *Am J Physiol Lung Cell Mol Physiol.* 2020;318:L533-L548
4. Karki P, Birukov KG, Birukova AA. Extracellular histones in lung dysfunction: A new biomarker and therapeutic target? *Pulm Circ.* 2020;10:2045894020965357
5. Karki P, Birukov KG. Oxidized phospholipids in healthy and diseased lung endothelium. *Cells.* 2020;9
6. Ke Y, Karki P, Kim J, Son S, Berdyshev E, Bochkov VN, Birukova AA, Birukov KG. Elevated truncated oxidized phospholipids as a factor exacerbating ALI in the aging lungs. *FASEB J.* 2019;33:3887-3900
7. Fang Y, Wu D, Birukov KG. Mechanosensing and mechanoregulation of endothelial cell functions. *Compr Physiol.* 2019;9:873-904
8. Karki P, Meliton A, Shah A, Tian Y, Ohmura T, Sarich N, Birukova AA, Birukov KG. Role of truncated oxidized phospholipids in acute endothelial barrier dysfunction caused by particulate matter. *PLoS One.* 2018;13:e0206251

Links:

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PubMed publications: <https://www.ncbi.nlm.nih.gov/pubmed/?term=birukov+k>

Wei Chao, MD, PhD, FAHA (wchao@som.umaryland.edu), Professor of Anesthesiology & Physiology. Dr. Chao is a Physician-Scientist and Co-Director of the Center for Shock, Trauma & Anesthesiology Research at the University of Maryland School of Medicine. His laboratory is interested in the molecular and cellular mechanisms of sepsis, traumatic injury, and ischemic myocardial injury, specifically the role of novel innate immune signaling in the pathogenesis of these critical illnesses. For these basic and translational studies, they use a combination of mouse genetics (transgenics and knockouts), physiology, biochemistry, and immunology. He has been PI on multiple grants from the NIH, DoD, and NSF. He has mentored both Ph.D. and M.D. scientists and his trainees have been successfully funded by various career development and independent research awards, such as NHLBI K08 Award and NIGMS ESI R35 Award.

Highlighted Publications:

1. Li Y, Feng Y, Chen H, Zou L, Si R, Wang E, Zhang M, Warren S, Sosnovik D, **Chao W**. Myocardial ischemia induces a rapid activation of innate immune signaling *via* cardiac heat-shock protein 60 and Toll-like receptor 4. *J Biol Chem*. 2011; 286:31308-19. PMID: 21775438
2. Chen C, Feng Y, Zou L, Chen HH, Cai JY, Xu JM, Sosnovik DE, **Chao W**. Role of extracellular RNA and TLR3-Trif signaling in myocardial ischemia-reperfusion injury. *J Am Heart Assoc*. 2014 Jan 3; 3(1): e000683. PMID: 24390148
3. Feng Y, Chen H, Cai J, Zou L, Yan D, Xu G, Li D, **Chao W**. Cardiac RNA induces inflammatory responses in cardiomyocytes and immune cells via Toll-like receptor 7 signaling. *J Biol Chem*. 2015; 290: 26688-98. PMID: 26363072.
4. Zou L, Feng Y, Xu G, Jian W, **Chao W**. Splenic RNA and microRNA mimics promote complement factor B production and alternative pathway activation via innate immune signaling. *J Immunol*. 2016; 196: 2788-98. PMID: 26889043.
5. Feng Y, Zou L, Yan D, Chen H, Xu G, Jian W, Cui P, **Chao W**. Extracellular microRNAs induce potent innate immune responses via TLR7/MyD88-dependent mechanisms. *J Immunol*. 2017; 199: 2106-17. PMID: 28768728.
6. Chen H., Yuan H., Cho H., Feng Y., Ngoy S., Kumar A., Liao R., **Chao W.**, Josephson L., Sosnovik D.. Theranostic Nucleic Acid Binding Nanoprobe Exerts Anti-inflammatory and Cytoprotective Effects in Ischemic Injury. *Theranostics*. 2017. 7(4), 814-825. PMID: 28382156
7. Xu J, Feng Y, Jeyaram A, Jay SM, Zou L, **Chao W**. Circulating plasma extracellular vesicles from septic mice induce inflammation via microRNA- and TLR7-dependent mechanisms. *J Immunol*. 2018; 201: 3392-3400. [PMID: 30355788](#)
8. Jian W, Gu L, Williams B, Feng Y, **Chao W** and Zou L. Toll-like Receptor 7 Contributes to Inflammation, Organ Injury, and Mortality in Murine Sepsis. *Anesthesiology*. 2019; 131:105-118; PMID: [31045897](#)
9. Williams B, Neder J, Cui P, Suen A, Tanaka K, Zou L and **Chao W**. Toll-like receptor 2 and 7 mediate coagulation activation and coagulopathy in murine sepsis. *J Thromb Haemost*. 2019; 17:1683-1693. [PMID: 31211901](#)
10. Jeyaram A, Lamichhane TN, Wang S, Zou L, Dahal E, Kronstadt SM, Levy D, Parajuli B, Knudsen DR, **Chao W**, Jay SM. Enhanced Loading of Functional miRNA Cargo via pH Gradient Modification of Extracellular Vesicles. *Mol Ther*. 2019 Dec 24 [Online ahead of print]. PMID: 31911034.
11. Chen F, Zou L, Williams B, and **Chao W**. Targeting Toll-Like receptors in sepsis - From bench to clinical trials. *Antioxid Redox Signal*. 2021; doi: 10.1089/ars.2021.0005. Online ahead of print. [PMID: 33588628](#)

Links:

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PubMed Publications:

<https://www.ncbi.nlm.nih.gov/sites/myncbi/1F3awzaP7DBkx/bibliography/47345926/public/?sort=date&direction=descending>

Christine Lau: Dr. Lau is Professor and Chair of the Department of Surgery at the University of Maryland, a thoracic surgeon specializing in lung transplantation, and a surgeon-scientist. Her research focuses on the pathogenesis and prevention of lung transplant rejection. Specifically, she has studied (1) biomarkers of bronchiolitis obliterans focusing on the CXCR4/CXCL12 axis,

NKT cells, and loss of epithelial cells; and (2) the therapeutic potential of adenosine analogs in mitigating lung transplant injury utilizing a porcine ex vivo lung perfusion and preclinical models.

Highlighted Publications:

1. Harris DA, Zhao Y, Lapar DL, Emaminia A, Steidle J, Kron IL, Lau CL. Inhibiting Fibrocyte Attenuates Bronchiolitis Obliterans in a Murine Tracheal Transplant Model, JTCVS, 2013 Mar;145(3):854-61.
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3. Gillen JR, Zhao Y, Harris DA, Lapar DJ, Stone ML, Fernandez LG, Kron IL, Lau CL. Rapamycin blocks fibrocyte migration and attenuates bronchiolitis obliterans in a murine model. Ann Thorac Surg. 2013 May;95(5):1768-75.
4. Lau CL, Zhao Y, Kron IL, Stoler MH, Laubach VE, Ailawadi G, Linden J. (2009) The role of adenosine A2A receptor signaling in bronchiolitis obliterans. Ann Thorac Surg. 88(4):1071-8.
5. Zhao Y, LaPar DJ, Steidle J, Emaminia A, Kron IL, Ailawadi G, Linden J, Lau CL. (2010) Adenosine signaling via the adenosine 2B receptor is involved in bronchiolitis obliterans development. J Heart Lung Transplant. 29:1405-14.
6. Emaminia A, Lapar DJ, Zhao Y, Steidle JF, Harris DA, Laubach VE, Linden J, Kron IL, Lau CL. Adenosine A(2A) Agonist Improves Lung Function During Ex Vivo Lung Perfusion. Ann Thorac Surg. 2011 Nov;92(5):1840-6.
7. Zhao Y, Steidle JF, Upchurch GR, Kron IL, Lau CL. Prevention of the second stage of epithelial loss is a potential novel treatment for bronchiolitis obliterans. J Thorac Cardiovasc Surg. 2013 Apr;145(4):940-947.
8. Gillen JR, Zhao Y, Harris DA, Lapar DJ, Kron IL, Lau CL. Short-course rapamycin treatment preserves airway epithelium and protects against bronchiolitis obliterans. Ann Thorac Surg. 2013 Aug;96(2):464-72.
9. Zhao Y, Gillen JR, Harris DA, Kron IL, Murphy MP, Lau CL. Treatment with placenta-derived mesenchymal stem cells mitigates development of bronchiolitis obliterans in a murine model. J Thorac Cardiovasc Surg. 2014 May;147(5):1668-1677
10. Stone ML; Sharma AK; Mas VR; Gehrau RC; Mulloy DP; Zhao Y; Lau CL; Kron IL; Huerter ME; Laubach VE. Ex Vivo Perfusion With Adenosine A2A Receptor Agonist Enhances Rehabilitation of Murine Donor Lungs After Circulatory Death. Transplantation. 99(12):2494-503, 2015.

Links:

Med School faculty page: <https://www.medschool.umaryland.edu/profiles/Lau-Christine/>